

New cysteine protease inhibitors in physiological secretory fluids and their medical significance

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Abstract: New cysteine protease inhibitors in human tears and milk were found and their medical significance was studied. As the protective components against bacterial infection in eyes, we detected four kinds of biologically active proteins in normal human tears including three kinds of cysteine protease inhibitors. Using our reverse zymography of normal tears, the three kinds of cysteine protease inhibitors were found to be 78, 20 and 15 kDa and were determined to be lactoferrin, VEG protein and cystatin S, respectively. The C-terminus area 17 mer peptide, Y₆₇₉-K₆₉₅ of lactoferrin molecule showed strong homology with a common active domain of cystatin family and the synthesized peptide itself showed considerable inhibition of cysteine proteases. Not only disease-specific changes of these inhibitor contents, but disease-specific new inhibitors were also found in patient tears in special autoimmune diseases. The characteristic 35 kDa inhibitor band which was detected specifically in the cases of Behcet's disease tears, an autoimmune disease, was determined to be a lachrymal acidic proline-rich protein family based on the N-terminus sequence analysis. The 65 kDa inhibitor of tears in Harada's autoimmune-disease was determined to be a human Ig heavy chain V-III region. Also lactoferrin content in Harada's disease was very low compared with that of normal tears. Also we found two cathepsin inhibitors, lactoferrin and κ -casein, in milk of human and bovine using reverse zymography. They may also play a role in bacterio-cidal and viro-cidal functions in milk. The L₁₃₃-Q₁₅₁ in human κ -casein molecule is the active inhibitory domain. It is most important to know from biological aspects that the concentration of these inhibitors in natural milk can inhibit cysteine proteases of bacteria. Surprisingly, the 50 times diluted milk inhibited papain completely, because lactoferrin and casein contents in milk are very high. We want to emphasize that these inhibitors in milk play a sufficient role in the protection of bacteria.

Index Keywords: casein; cathepsin; cystatin; cysteine proteinase inhibitor; lactoferrin; papain; peptide; transferrin; amino acid sequence; animal; breast milk; cattle; chemistry; conference paper; dose response; human; kinetics; liver; metabolism; milk; molecular genetics; polyacrylamide gel electrophoresis; protein tertiary structure; rat; sequence homology; structure activity relation; Amino Acid Sequence; Animals; Caseins; Cathepsins; Cattle; Cystatins; Cysteine Proteinase Inhibitors; Dose-Response Relationship, Drug; Electrophoresis, Polyacrylamide Gel; Humans; Kinetics; Lactoferrin; Liver; Milk; Milk, Human; Molecular Sequence Data; Papain; Peptides; Protein Structure, Tertiary; Rats; Sequence Homology, Amino Acid; Structure-Activity Relationship; Transferrin; Animalia; Bacteria (microorganisms); Bos taurus; Bovinae

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References:

1. Barrett, A.J., Kirschke, H., Cathepsin B, cathepsin H, cathepsin L (1981) *Methods Enzymol*, 80, pp. 535-561
2. Bhimani, R.S., Vendrov, Y., Furmanski, P., Influence of lactoferrin feeding and injection against systemic staphylococcal infections in mice (1999) *J Appl Microbiol*, 86 (1), pp. 135-144
3. Bossard, M.J., Tomaszek, T.A., Thompson, S.K., Amegadzie, B.Y., Hanning, C.R., Jones, C., Kurdyla, J.T., Levy, M.A., Proteolytic activity of human osteoclast cathepsin K. Expression, purification, activation, and substrate identification (1996) *J Biol Chem*, 271, pp. 12517-12524
4. Fernandez, C.P., Castellanos, S., Rodriguez, P., Reverse staining of SDS-PAGE gels by imidazole-zinc salt. Sensitive detection of unmodified protein (1992) *Biotechniques*, 12, pp. 564-573

5. Hof, W.V., Blankenvoorde, M.F., Veerman, E.C., Amerongen, A.V., The salivary lipocalin von Ebner's gland protein is a cysteine proteinase inhibitor (1997) *J Biol Chem*, 272 (3), pp. 1837-1841
6. Isemura, S., Saitoh, E., Ito, S., Isemura, M., Sanada, K., Cystatin, S., A cysteine proteinase inhibitor of human saliva (1984) *J Biochem*, 96, pp. 1311-1314
7. Katunuma, N., Kominami, E., Structure, properties, mechanisms, and assays of cysteine protease inhibitors: Cystatins and E-64 derivatives (1995) *Methods Enzymol*, 251, pp. 382-397
8. Katunuma, N., Matsunaga, Y., Saibara, T., Mechanism and regulation of antigen processing by cathepsin B (1994) *Adv Enzyme Regul*, 34, pp. 145-158
9. Katunuma, N., Murata, E., Kakegawa, H., Matsui, A., Tsuzuki, H., Tsuge, H., Turk, D., Asao, T., Structure based development of novel specific inhibitors for cathepsin L and cathepsin S in vitro and in vivo (1999) *FEBS Lett*, 458, pp. 6-10
10. Kopitar, G., Dolinar, M., Strukelj, B., Pungercar, J., Turk, V., Folding and activation of human procathepsin S from inclusion bodies produced in *Escherichia coli* (1996) *Eur J Biochem*, 236, pp. 558-562
11. Korant, B.D., Brzin, J., Turk, V., Cystatin, a protein inhibitor of cysteine proteases alters viral protein cleavages in infected human cells (1985) *Biochem Biophys Res Commun*, 127, pp. 1072-1076
12. Laemmli, K.U., Cleavage of structural protein during the assembly of the head of bacteriophage T4 (1970) *Nature*, 227, pp. 680-685
13. Maekawa, Y., Himeno, K., Ishikawa, H., Hisaeda, H., Sakai, T., Dainichi, T., Asao, T., Katunuma, N., Switch of CD4+ T Cell differentiation from Th2 to Th1 by treatment with cathepsin B inhibitor in experimental Leishmaniasis (1998) *J Immunol*, 161, pp. 2120-2127
14. Majima, E., Ishida, M., Miki, S., Shimohara, Y., Tada, H., Specific labeling of the bovine mitochondrial phosphate carrier with fluorescein 5-isothiocyanate (2001) *J Biol Chem*, 276 (13), pp. 9792-9799
15. Matsunaga, Y., Saibara, T., Kido, H., Katunuma, N., Participation of cathepsin B in processing of antigen presentation to MHC class II (1993) *FEBS Lett*, 324 (3), pp. 325-330
16. Nagaya, T., Murata, Y., Yamaguchi, S., Nomura, Y., Ohmori, S., Fujieda, M., Katunuma, N., Seo, H., Intracellular proteolytic cleavage of 9-cis-retinoic acid receptor? by cathepsin L-type protease is a potential mechanism for modulating thyroid hormone action (1998) *J Biol Chem*, 273 (50), pp. 33166-33173
17. Shimizu, N., Ikehara, Y., Tatematsu, M., (2002) Effect of Lactoferrin in H. Pylori Infection Animal Model. Lactoferrin, pp. 209-251. , Amsterdam: Elsevier
18. Takahashi, M., Tezuka, T., Katunuma, N., Inhibition of growth and cysteine proteinase activity of *Staphylococcus aureus* V8 by phosphorylated cystatin ? in skin cornified envelop (1994) *FEBS Lett*, 355, pp. 275-278
19. Zhang, T., Maekawa, Y., Hanba, J., Dainichi, T., Nashed, B.F., Hisaeda, H., Sakai, T., Katunuma, N., Lysosomal cathepsin B plays an important role in antigen processing, while cathepsin D is involved in degradation of the invariant chain in ovalbumin-immunized mice (2000) *Immunology*, 100, pp. 13-20

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